

# FUCHSIN ACID VAN GIESON REAGENT

IVD *In vitro* diagnostic medical device

CE

**For use with Van Gieson Trichrome and Weigert-Van Gieson kits, synonym Picrofuchsin reagent**

## INSTRUCTIONS FOR USE

REF Catalogue number: FAG-OT-100 (100 mL)

### Introduction

Fuchsin Acid Van Gieson kit is used for staining collagen, muscle tissue, keratinized epithelium, cytoplasm, glial fibers, and erythrocytes. The reagent contains two dyes (acid fuchsin, picric acid) that simultaneously stain different tissue structures. Acid fuchsin stains collagen fibers intensive red while picric acid stains muscle fibers, erythrocytes and glial fibers yellow. Amyloids, hyalin, colloid and mucosa are stained in nuances between red and yellow. Fuchsin Acid Van Gieson reagent is a component of Van Gieson Trichrome kit, as well as Weigert-Van Gieson kits.

### Product description

- **FUCHSIN ACID VAN GIESON REAGENT** – reagent containing Fuchsin Acid dye and picric acid for selective staining of tissue structures

### Example of Fuchsin Acid Vane Gieson reagent usage as a component of Van Gieson Trichrome kit

#### Other sections and reagents that may be used in staining:

- Fixatives such as BioGnost's neutral buffered formaldehyde solutions: Formaldehyde NB 4%, Formaldehyde NB 10%
- Dehydrating/rehydrating agent, such as BioGnost's alcohol solutions: Histanol 70, Histanol 80, Histanol 95 and Histanol 100
- Clearing agents, such as BioClear xylene or a substitute, such as BioClear New agent on the aliphatic hydrocarbons basis
- Infiltration and fitting agent, such as BioGnost's granulated paraffin BioWax Plus, BioWax 56/68, BioWax Blue, BioWax Micro.
- High-quality glass slides for use in histopathology and cytology, such as VitroGnost SUPER GRADE, VitroGnost COLOR or one of more than 30 models of BioGnost's VitroGnost glass slides
- BioGnost's reagents that comprise Hematoxylin Weigert: Hematoxylin, Weigert A and Ferri reagent, Weigert B

### Preparation of additional solutions used in staining

#### Hematoxylin Weigert solution

- Mix Hematoxylin, Weigert A and Ferri reagent, Weigert B in 1:1 ratio. Hematoxylin Weigert solution is stable for approximately 2 weeks. Discard the solution when the nuclei turn brown after staining.

#### Preparing the histological sections for staining

- Fixate the sample (Formaldehyde NB 4%, Formaldehyde NB 10%), rinse with water and dehydrate through series of ascending alcohol solutions (Histanol 70, Histanol 80, Histanol 95 and Histanol 100).
- Clear the sample with intermedium; in xylene (BioClear) or in a xylene substitute (BioClear New).
- Infiltrate and fit the sample in paraffin (BioWax Plus, BioWax 56/58, BioWax Blue, BioWax Micro).
- Cut the paraffin block to 4-6  $\mu\text{m}$  slices and place them on a VitroGnost glass slide.

### Procedure of staining histology samples by using Van Gieson Trichrome kit with three 100 ml reagents (VG-K-100)

Pour Fuchsin Van Gieson reagent into glass staining jars (Coplin, Hellendahl or Schifferdecker), return to original bottles after staining. Close tightly. Filter the reagent if necessary.

1.	Deparaffinize the section in xylene (BioClear) or in a xylene substitute (BioClear New)	3 exchanges, 2 min each
2.	Rehydrate using 100% alcohol (Histanol 100)	2 exchanges, 5 and 3 min
3.	Rehydrate using 95% alcohol (Histanol 95)	2 min
4.	Rehydrate in distilled (demi) water	2 min
5.	Immerse into Hematoxylin Weigert solution	5 min
6.	Rinse under tap water	3 min
7.	Immerse into Fuchsin Acid Van Gieson reagent	30 seconds
	Note: Exposition longer than 30 seconds causes the sections to turn intensive red.	
8.	Rinse in distilled (demi) water	30 seconds
9.	Dehydrate using 95% alcohol (Histanol 95)	2 exchanges, 30 seconds each
10.	Dehydrate using 100% alcohol (Histanol 100)	2 exchanges, 1 min each
11.	Clear the section in xylene (BioClear) or in a xylene substitute (BioClear New)	2 exchanges, 2 min each

Immediately after clearing apply an appropriate BioMount medium for covering/mounting on the section. If BioClear xylene was used, use one of BioGnost's mounting xylene-based media (BioMount, BioMount High, BioMount M, BioMount DPX, BioMount C, or universal BioMount New). If BioClear New xylene substitute was used, the appropriate covering agent is BioMount New. Cover the section with a VitroGnost cover glass.

### Note

Time periods of staining processes are not entirely standardized and they approximately correspond to clinical and laboratory practical experience. Intensity of staining depends on the period of immersion in the dye. Real staining protocol depends on personal requests and priorities.

## Result

Blue-brown - nuclei

Red - collagen

Yellow - muscle fiber, glial fibers

Hues between yellow and red - colloid, mucosa, hyalin, amyloid, keratinized epithelium

## Preparing the sample and diagnostics

Use only appropriate instruments for collecting and preparing the samples. Process the samples with modern technology and mark them clearly. Follow the manufacturer's instructions for handling. In order to avoid mistakes, the staining procedure and diagnostics should only be conducted by authorized and qualified personnel. Use only microscope according to standards of the medical diagnostic laboratory.

## Safety at work and environmental protection

Handle the product in accordance with safety at work and environmental protection guidelines. Used solutions and out of date solutions should be disposed of as special waste in accordance with national guidelines. Chemicals used in this procedure could pose danger to human health. Tested tissue specimens are potentially infectious. Necessary safety measures for protecting human health should be taken in accordance with good laboratory practice. Act in accordance with signs and warnings notices printed on the product's label, as well as in BioGnost's material safety data sheet.














## Storing, stability and expiry date

Keep Fuchsin Acid Van Gieson reagent in a tightly closed original package at temperature between 15°C and 25°C. Do not freeze and avoid exposing to direct sunlight. Date of manufacture and expiry date are printed on the product's label.

## References

1. Culling, C.F.A. (1974): Handbook of histopathological and histochemical techniques, 2<sup>nd</sup> ed., Butterworth, London, UK.
2. Lillie, R.D. (1945): Studies on selective staining of collagen with acid aniline dyes, J. Technical Methods, 25:1
3. Sheehan D.C. et Hrapchak, B.B. (1980): Theory and Practice Histotechnology, 2<sup>nd</sup> ed., CV Mosby, St. Louis, (MO), pp 52, p 14-167.
4. Van Gieson, I. (1889): Laboratory notes of technical methods for the nervous system, New York Med. J., 50: 57-60

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	Refer to the supplied documentation		Storage temperature range		Number of tests in package		Product code		European Conformity
	Refer to supplied instructions		Keep away from heat and sunlight		Valid until		Lot number		Manufacturer
	For <i>in vitro</i> diagnostic use only		Keep in dry place		Caution - fragile				



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