

OG-EA50 PAP REAGENT

IVD *In vitro* diagnostic medical device



Rapid staining reagent acc. to Papanicolaou Counterstain for monochromatic staining of samples in cytology

INSTRUCTIONS FOR USE

REF Catalogue number: OGEA50-OT-100 (100 mL) OGEA50-OT-500 (500 mL) OGEA50-OT-1L (1000 mL) OGEA50-OT-2.5L (2500 mL)

Introduction

OG-EA50 Pap reagent is alcoholic polychromatic solution. The first step in using the Papanicolaou staining method implies nuclear staining with a hematoxylin solution, and next two steps consist of counterstaining using the monochromatic OG-6 reagent and one of the polychromatic EA reagent formulations. OG-EA50 Pap reagent does not require three, but two steps because it contains OG-6 and EA50 reagents. The Orange G molecule stains the cytoplasm, and in later stages of the procedure it remains only in the mature, keratinized cells. Polychromatic EA50 solution stains non-stained parts of cell, such as squamous cells, nucleoli, ciliary and erythrocytes. Test samples can be gynecological and non-gynecological, such as sputum, urine, and cytological puncture samples. In order to obtain optimal staining results, OG-EA50 Pap reagent has properties completely in compliance with other BioGnost's reagents for cytological smearing acc. to Papanicolaou - Hematoxylin HP, Pap 1A reagent and Hematoxylin HP, Pap 1B reagent.

Product description

OG-EA50 PAP REAGENT - Counterstain for polychromatic staining of gynecology samples in cytology. Enables rapid staining reagent acc. to Papanicolaou

Preparing the cytological smear for staining

There are two methods of collecting and preparing the cytological samples:

1. After collecting the cytological sample, place it on the microscope slide (VitroGnost), fixate it immediately with a fixative in a spray bottle (CitoSpray), dry it and keep until the staining process. Cytological sample may be fixated and kept until staining by immersing into 95% alcohol solution (Histanol 95) for a minimum of 30 minutes.
2. Using liquid-based cytology method (LBC) and brush for collecting cytological samples, fixate the sample immediately (CitoFix, CitoFix in transport containers) by removing the brush head and immersing it in the fixative. At the beginning of processing the sample, isolate the cells from the fixative (one of the methods is to centrifuge the fixative) and place them on the microscope slide equally in a single layer. Cytological sample prepared in such a way is ready for staining.

The Papanicolaou staining method, **PROGRESSIVE**

The first stage of staining procedure depends on the method the cytological sample was collected and fixated on the microscope slide.

If the sample is dry and previously fixed using CitoSpray, it is necessary to keep it in a 95% alcohol solution (Histanol 95) for 10 minutes in order to remove polyglycols. If the section was fixated with a 95% alcohol solution (Histanol 95), ignore this step. During staining cytology samples (prepared by using the liquid based cytology method (LBC)) that contain low concentration of alcohol, rehydration by descending series of alcohol solutions is not necessary. The procedure starts by rinsing the section using distilled (demi) water and is then stained using Hematoxylin HP, Pap 1A reagent.

1.	Rehydrate in descending series of alcohols (Histanol 95, Histanol 80 and Histanol 70) and in distilled or demineralized water	6-8 dips in each of the 4 exchanges
2.	Stain using Hematoxylin HP, Pap 1A reagent	30 seconds
3.	Rinse the section with tap or distilled water	10 seconds
4.	Blue using Scott's solution or Bluing reagent	1 min
	Note: If the mentioned reagents are not available, the section should be blued using indirect stream of water	3-5 minutes
5.	Immerse the sections in distilled/demineralized water.	10 seconds
6.	Dehydrate in ascending series of alcohols (Histanol 70, Histanol 80 and Histanol 95)	6-8 dips in each of the 3 exchanges
7.	Stain using OG-EA50 Pap reagent	4 min
8.	Rinse using 95% alcohol in two exchanges (Histanol 95)	6-8 dips in each of the 2 exchanges
9.	Dehydrate using 100% alcohol (Histanol 100)	6-8 dips
10.	Dehydrate using 100% alcohol (Histanol 100)	3-5 minutes
11.	Clear the section in xylene (BioClear) or in a xylene substitute (BioClear New)	6-8 dips
12.	Clear the section in xylene (BioClear) or in a xylene substitute (BioClear New)	3-5 minutes

Immediately after clearing apply an appropriate BioMount medium for covering/mounting on the section. If BioClear xylene was used, use one of BioGnost's mounting xylene-based media (BioMount, BioMount High, BioMount M, BioMount DPX, BioMount C, or universal BioMount New). If BioClear New xylene substitute was used, the appropriate covering agent is BioMount New. Cover the section with VitroGnost cover glass.

Note

In the case of subsidence in the Hematoxylin HP, Pap 1A solution or formation of metallic glow on the surface, reagent should be filtered before use. Time periods of staining procedures are not completely standardized. The suggested methods are in accordance with BioGnost's reagents' properties and correspond to longtime clinical and laboratory practice. Intensity of staining depends on the period of exposure to stains and reagents. Staining procedure can be changed according to personal preferences if they correspond to the basic principles of cytotechnology.

Papanicolaou staining method, REGRESSIVE

The regressive staining method creates a better sample differentiation and clearer nuclear structure visibility.

The first stage of staining procedure depends on the method the cytological sample was collected and fixated on the microscope slide.

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1.	Rehydrate in descending series of alcohols (Histanol 95, Histanol 80 and Histanol 70) and in distilled or demineralized water	6-8 dips in each of the 4 exchanges
2.	Stain using Hematoxylin HP, Pap 1A reagent	3 min
3.	Rinse the section with tap or distilled water	30 seconds
4.	Differentiate using HCL Pap reagent or in 0.1% HCl solution	10 seconds
5.	Rinse the section with tap or distilled water	30 seconds
6.	Blue using Scott's solution or Bluing reagent	1 min
	Note: If the mentioned reagents are not available, the section should be blued using indirect stream of water	3-5 minutes
7.	Immerse the sections in distilled/demineralized water.	10 seconds
8.	Dehydrate in ascending series of alcohols (Histanol 70, Histanol 80 and Histanol 95)	6-8 dips in each of the 3 exchanges
9.	Stain using OG-EA50 Pap reagent	5 min
10.	Stain using 95% alcohol in two exchanges (Histanol 95)	6-8 dips in each of the 2 exchanges
11.	Dehydrate using 100% alcohol (Histanol 100)	6-8 dips
12.	Dehydrate using 100% alcohol (Histanol 100)	3-5 minutes
13.	Clear the section in xylene (BioClear) or in a xylene substitute (BioClear New)	6-8 dips
14.	Clear the section in xylene (BioClear) or in a xylene substitute (BioClear New)	3-5 minutes

Immediately after clearing apply an appropriate BioMount medium for covering/mounting on the section. If BioClear xylene was used, use one of BioGnost's mounting xylene-based media (BioMount, BioMount High, BioMount M, BioMount DPX, BioMount C, or universal BioMount New). If BioClear New xylene substitute was used, the appropriate covering agent is BioMount New. Cover the section with VitroGnost cover glass.

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Results

Blue - nuclei

Yellow-orange - keratinized cells

Pink-red - superficial squamous epithelial cell, erythrocytes, nucleoli, cilia

Green - cytoplasm of all the other cell types (parabasal and intermediate squamous cells, columnar cells, polymorphonuclear leukocytes, lymphocytes, histiocytes, adenocarcinomas, undifferentiated carcinoma cells)

Preparing the sample and diagnostics

Use only appropriate instruments for collecting and preparing the samples. Process the samples with modern technology and mark them clearly. Follow the manufacturer's instructions for handling. In order to avoid mistakes, the staining procedure and diagnostics should only be conducted by authorized and qualified personnel. Use only microscope according to standards of the medical diagnostic laboratory.

Safety at work and environmental protection

Handle the product in accordance with safety at work and environmental protection guidelines. Used solutions and out of date solutions should be disposed of as special waste in accordance with national guidelines. Chemicals used in this procedure could pose danger to human health. Tested tissue specimens are potentially infectious. Necessary safety measures for protecting human health should be taken in accordance with good laboratory practice. Act in accordance with signs and warnings notices printed on the product's label, as well as in BioGnost's material safety data sheet.














Storing, stability and expiry date

Keep OG-EA50 Pap reagent in a tightly closed original package at temperature between 15°C and 25°C. Keep in dry places, do not freeze and avoid exposing to direct sunlight. Date of manufacture and expiry date are printed on the product's label.

References

1. Carson, F.L., Hladik C. (2009): Histotechnology: A self-instructional text, 3rd ed. ASCP Press.
2. Papanicolaou, G.N. (1941): Some improved methods for staining vaginal smears. J Lab Clin Med.
3. Papanicolaou, G.N. (1942): A new procedure for staining vaginal smears. Science.
4. Sherwani, R.K., Khaq, T. et al. (2007): Conventional Pap Smear and Liquid Based Cytology for Cervical Cancer Screening – A Comparative Study, Journal of Cytology, 24 (4): pp 167-172.

OG-EA50-OT-X, V1-EN1, 22 September 2017, IK/AK

 Refer to the supplied documentation	 Storage temperature range	 Number of tests in package	 Product code	 European Conformity
 Refer to supplied instructions	 Keep away from heat and sunlight	 Valid until	 Lot number	 Manufacturer
 For <i>in vitro</i> diagnostic use only	 Keep in dry place	 Caution - fragile		

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