

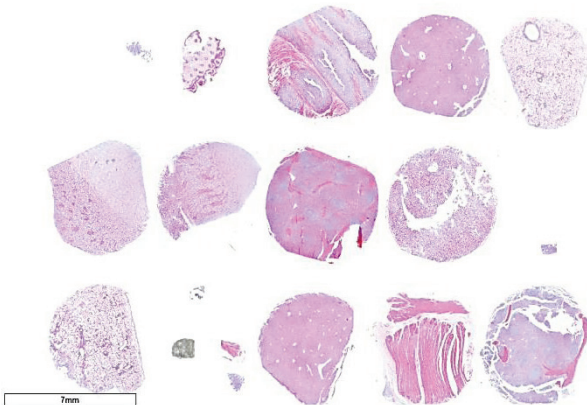
Hematoxylin substitute **ErioGnost reagent**

ErioGnost reagent is used in histology as an economic and ecologically acceptable synthetic replacement for hematoxylin. Specially formulated with optimal ratio of active dye and mordant, ErioGnost reagent demonstrates a superior stability compared to hematoxylin solutions. The staining method is compatible with conventional eosin counterstaining as a substitute for hematoxylin – eosin staining. It can also be used in special staining kits (such as A.F.O.G., Gomori Trichrome, Masson-Goldner Trichrome, Masson Trichrome, Picro Sirius Red, Safranin O and Mucicarmine) and as a counterstain in immunohistochemical staining methods (IHC).

COMPARISON OF STAINING METHODS*:

ErioGnost – eosin staining

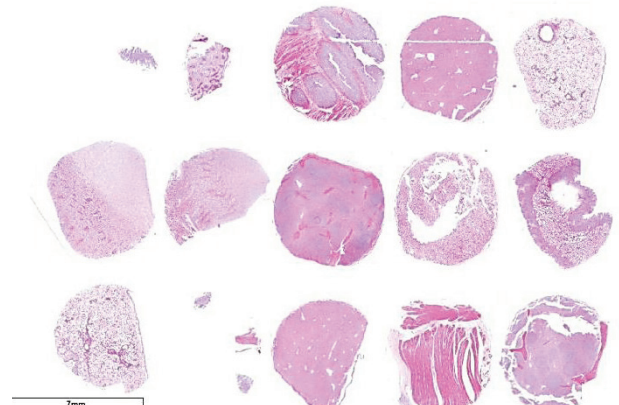
Nuclei – blue to purple
Cytoplasm, muscle/connective tissue – light pink to purple
Red erythrocytes



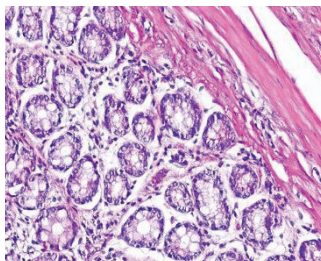
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Hematoxylin – eosin staining

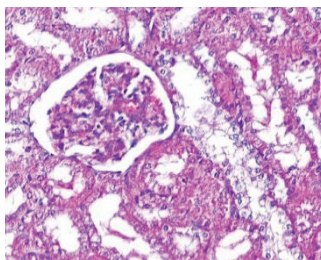
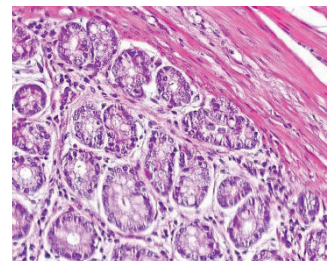
Nuclei - purple
Cytoplasm, muscle/connective tissue – dark pink to purple
Pink erythrocytes



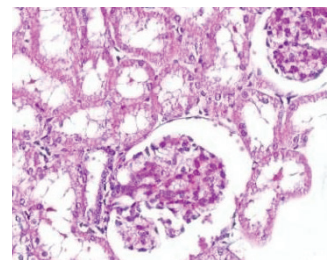
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Colon

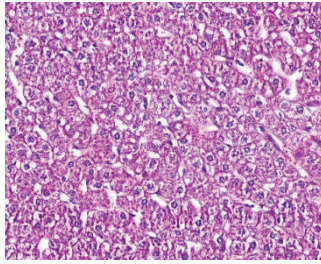


Kidney

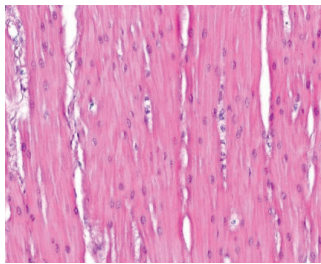
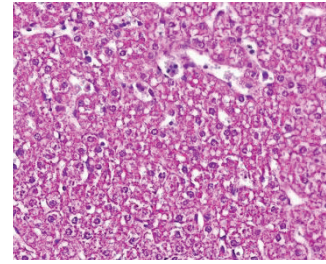


ErioGnost – eosin staining

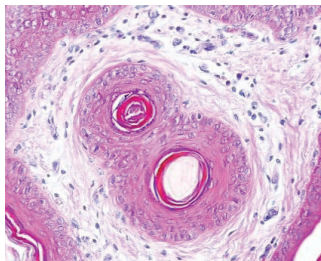
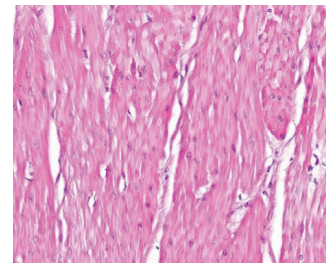
Hematoxylin – eosin staining



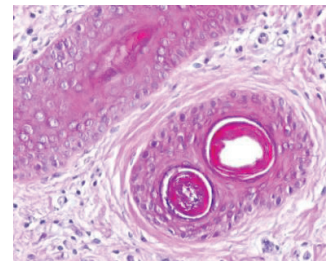
Liver



Muscle tissue



Skin



*Staining procedure:

(reagents used: BioGnost Harris hematoxylin; Hematoxylin H; product code HEMH-OT-100, HEMH-OT-500, HEMH-OT-1L, HEMH-OT-2.5L, Eosin Y 2% aqueous (product code EOYA-20-OT-1L, EOYA-20-OT-2.5L))

1. Deparaffinize the section in xylene (BioClear) or in a xylene substitute (BioClear New)	3 exchanges, 2 min each
2. Rehydrate using 100% alcohol (Histanol 100)	2 exchanges, 5 and 3 min
3. Rehydrate using 95% alcohol (Histanol 95)	2 min
4. Rehydrate in distilled (demi) water	2 min
5. Stain with ErioGnost reagent / Hematoxylin H	5 min / 4 min
6. Rinse under tap water until the dye is no longer released from the section	2 exchanges, 15 seconds each if ErioGnost reagent was applied / 3 minute rinsing under tap water if Hematoxylin H was applied
7. If alcoholic eosin solution is used, immerse the sections in 95% alcohol (Histanol 95). If aqueous eosin solution is used, skip this step.	
8. Stain using an eosin counterstain solution until section is stained optimally Note: Intensive eosinophilic dye is achieved much quicker by staining the sections in alcoholic eosin solutions (within 15 seconds); sections should be exposed to aqueous eosin solutions for 90 seconds to 2 minutes	15 seconds - 2 min
9. Rinse under tap water Note: If alcoholic eosin solution is used as counterstain, skip this step.	2 min
10. Dehydrate in 95% alcohol (Histanol 95)	2 exchanges, 10-15 dips each
11. Dehydrate u 100% alcohol (Histanol 100)	3 exchanges, 10-15 dips each
12. Clear the section in xylene (BioClear) or in a xylene substitute (BioClear New)	2 exchanges, 2 min each

